

NEW GUAIANOLIDES FROM *PSEUDOSTIFFTIA KINGII**

FERDINAND BOHLMANN, PAHUP SINGH, ROBERT M. KING† and HAROLD ROBINSON

Institute for Organic Chemistry, Technical University of Berlin, D-1000 Berlin 12, West Germany; †Smithsonian Institution, Department of Botany, Stop No. 166, Washington, DC 205060, U.S.A.

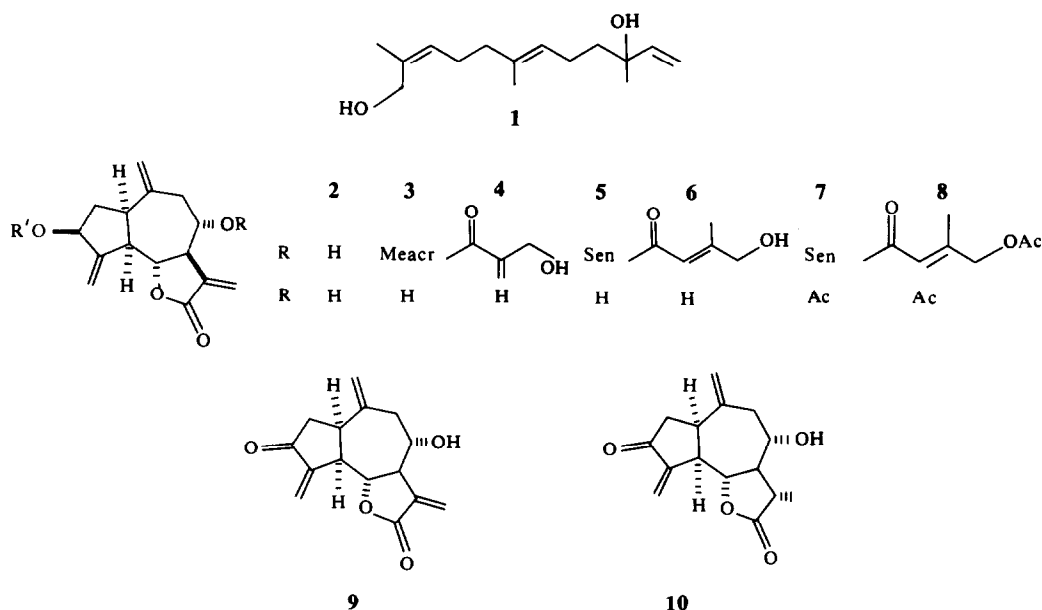
(Received 14 August 1981)

Key Word Index—*Pseudostiffia kingii*; Compositae; Vernoniaeae; sesquiterpene lactones; guaianolides.

Abstract—*Pseudostiffia kingii* afforded several guaianolides, two of them not isolated before.

Pseudostiffia is a monotypic genus and the single genus in the subtribe Pseudostiffinae[1], where it has been placed with reservation as the thickened style with scabrid surface and the large non-lophorate pollen grains are unlike any other member of the Vernoniaeae, although it also does not fit within any other tribe. A first chemical investigation only afforded acetylenes and triterpenes[2]. We now have re-investigated the aerial parts of *Pseudostiffia kingii* H. Robins. In addition to the compounds previously

guaianolides desacylcynaropicrin (2)[4], arguerin B (3)[5], cynaropicrin (4)[6], 8 α -hydroxy-dehydrozaluzanin C (9)[7], the corresponding 11 β ,13-dihydro derivative 10[8] as well as two further lactones, the senecioate 5 and the hydroxysenecioate 6. The structures followed from the ¹H NMR spectra (Table 1). The data of 5 clearly showed the presence of a zaluzanin C derivative, as most signals were similar to those of zaluzanin C. However, an additional seneciyoxy residue was present. Spin



isolated and 13-hydroxynorolidol (1)[3], minute amounts of sesquiterpene lactones were isolated, the

decoupling showed that this group had to be placed at C-8, while from the couplings the α -orientation could be deduced. Acetylation afforded the acetate 7, its ¹H NMR spectral data further supporting the proposed structure. The ¹H NMR spectral data of 6 indicated that a hydroxylated ester group was present, obviously the unusual 4-hydroxysenecioate. Since the signal of the olefinic methyl group was at δ 2.15, the

*Part 414 in the series "Naturally Occurring Terpene Derivatives". For Part 413 see Bohlmann, F., Singh, P., Ruke, A., King, R. M. and Robinson, H. (1982) *Phytochemistry* 21 (in press).

Table 1. ^1H NMR spectral data of compounds 5–8 (400 MHz, CDCl_3 , TMS as int. standard)

	5	6	7	8
H-1	2.98 dddd	3.02 dddd	2.98 dddd	3.02 dddd
H-2 α	1.74 ddd	1.80 ddd	1.75 ddd	1.80 ddd
H-2 β	2.24 ddd	2.39 ddd	2.24 ddd	2.38 ddd
H-3	4.56 br dd	5.57 dddd	4.55 br dd	5.57 dddd
H-5	2.85 br dd	2.85 br dd	2.85 br dd	2.85 br dd
H-6	4.22 dd	4.17 dd	4.22 dd	4.17 dd
H-7	3.12 dddd	3.13 dddd	3.13 dddd	3.15 dddd
H-8	5.04 ddd	5.05 ddd	5.06 ddd	5.09 ddd
H-9 α	2.70 dd	2.70 dd	2.70 dd	2.70 dd
H-9 β	2.36 dd	2.39 dd	2.37 dd	2.38 dd
H-13	6.22 d	6.23 d	6.22 d	6.24 d
H-13'	5.66 d	5.67 d	5.66 d	5.65 d
H-14	5.14 br s	5.13 br s	5.14 br s	5.14 br s
H-14'	4.96 br s	4.98 br s	4.97 br s	4.99 br s
H-15	5.51 ddd	5.53 dd	5.50 dd	5.54 dd
H-15'	5.38 ddd	5.35 dd	5.36 dd	5.36 dd
OCOR	5.74 qq	5.74 qq	6.08 q	5.92 q
	2.20 d	2.20 d	4.20 br s	4.62 br s
	1.95 d	1.95 d	2.15 br s	2.20 br s
OAc	—	2.10 s	—	2.17 s
				2.11 s

$J(\text{Hz})$: 1,2 α = 11; 1,2 β = 1,5 = 8; 2 α ,2 β = 13; 2 α ,3 = 2 β ,3 = 8; 3,15 = 5,15 = 1.5; 5,6 = 10.5; 6,7 = 9; 7,8 = 10; 7,13 = 3.5; 7,13' = 3; 8,9 α = 5; 8,9 β = 4; 9 α ,9 β = 15.

stereochemistry was also clear. All other signals were similar to those of 5. Acetylation afforded the diacetate 8, its ^1H NMR spectrum further establishing the structure. Finally a trace of a further lactone was isolated, which obviously was a furanoheliangolide with an angelate residue, deduced from the typical ^1H NMR signals of H-2, H-5, H-14, H-15 and angelate. However, the amount of material was not sufficient to establish the structure.

These additional results on the chemistry of *Pseudostiffia* support the placement of this genus in the Vernoniaeae, since guaianolides and lupanes are widespread in the tribe.

EXPERIMENTAL

The air dried aerial parts (100 g), collected in north-eastern Brazil, voucher RMK 8666, deposited in the U.S. National Herbarium, Washington, was extracted with Et_2O -petrol, 1:2 and the resulting extract was separated by CC (Si gel) and further by repeated TLC (Si gel). Known compounds were identified by comparing the ^1H NMR spectra with those of authentic material. In addition to compounds isolated before, 2 mg 1, 5 mg 2, 1.5 mg 3, 4 mg 4, 2 mg 5 (Et_2O -petrol, 3:1), 2.5 mg 6 (Et_2O), 5 mg 9 and 0.5 mg 10 were obtained.

8 α -(4'-Hydroxyseneciolyloxy)-dehydrozaluazinin C (5). Colourless gum, not free from 3, IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm^{-1} : 3600 (OH), 1775 (γ -lactone), 1720 ($\text{C}=\text{CCO}_2\text{R}$); MS m/z (rel. int.): 344.164 $[\text{M}]^+$ (1) ($\text{C}_{20}\text{H}_{24}\text{O}_5$), 244 $[\text{M}-\text{HO}_2\text{CR}]^+$ (23), 226 $[244-\text{H}_2\text{O}]^+$ (12), 83 $[\text{C}_4\text{H}_7\text{CO}]^+$ (100), 55 $[83-\text{CO}]^+$ (40).

$$[\alpha]_D^{25} = \frac{589}{+32} \frac{578}{+33} \frac{546}{+37} \frac{436 \text{ nm}}{+68} (\text{CHCl}_3; c 0.2).$$

Acetylation (1 hr, Ac_2O , 70°) afforded 7, a colourless gum; ^1H NMR see Table 1.

8 α -(4'-Hydroxyseneciolyloxy)-dehydrozaluazinin C (6). Colourless gum, which was purified as its acetate 8 also a colourless gum, IR $\nu_{\text{max}}^{\text{CCl}_4}$ cm^{-1} : 3605 (OH), 1780 (γ -lactone), 1740 (OAc), 1720 ($\text{C}=\text{CCO}_2\text{R}$); MS m/z (rel. int.): 444 $[\text{M}]^+$ (0.1), 286 $[\text{M}-\text{HO}_2\text{CR}]^+$ (0.2), 226.099 $[286-\text{HOAc}]^+$ (9) ($\text{C}_{15}\text{H}_{14}\text{O}_2$), 211 $[226-\text{Me}]^+$ (1), 198 $[226-\text{CO}]^+$ (3), 141 $[\text{RCO}]^+$ (7), 99 $[\text{C}_4\text{H}_6(\text{OH})\text{CO}]^+$ (100).

$$[\alpha]_D^{25} = \frac{589}{+22} \frac{578}{+24} \frac{546}{+36} \frac{436 \text{ nm}}{+98} (\text{CHCl}_3; c 0.25).$$

Acknowledgements—We thank Drs. Scott A. Mori and P. Alvim, Herbario Centro de Pesquisas do Cacau at Itabanu, Bahia, Brazil, for their help during plant collection and the Deutsche Forschungsgemeinschaft for financial support.

REFERENCES

1. Robinson, H., Bohlmann, F. and King, R. M. (1980) *Phytologia* **46**, 421.
2. Bohlmann, F., Zdero, C., King, R. M. and Robinson, H. (1980) *Phytochemistry* **19**, 2669.
3. Bohlmann, F. and Zdero, C. (1980) *Phytochemistry* **19**, 587.
4. Gonzales, A. G., Bermejo, J., Massanet, G. M. and Perez, J. (1973) *An. Quim.* **1333**.
5. Gonzales, A. G., Bermejo, J., Cabrera, I., Massanet, G. M., Mansilla, H. and Galindo, A. (1978) *Phytochemistry* **17**, 955.
6. Suchy, M., Herout, V. and Sorm, F. (1959) *Collect. Czech. Chem. Commun.* **24**, 1542.
7. Bohlmann, F., Singh, P., Borthakur, N. and Jakupovic, J. (1981) *Phytochemistry* **20**, 2379.
8. Bohlmann, F., Brindöpke, G. and Rastogi, R. C. (1978) *Phytochemistry* **17**, 475.